

PHENOL TESTS

I. A CLASSIFICATION OF THE TESTS AND A REVIEW OF THE LITERATURE

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INTRODUCTION

Chemists and biologists have found extensive use for tests of phenolic compounds. Compounds of this class are not only widely employed in industry but are distributed in liberal variety throughout the animal and vegetable kingdoms. Many are found by the biologists to occur naturally as products of metabolism; several are used as medicinals or antiseptics.

Their varied importance led to many methods of detection; and, since quantities available were often small, there has been continual effort to develop delicate color tests and wherever possible to make the tests specific. More than 40 of the tests vary in the character of the reagents, and these have been modified, improved and complicated in various ways so that now the tests described in the literature number more than 100.

It is of interest to note that while these tests are designed to detect phenols, very few of the reactions employed involve the hydroxyl group directly. They are brought about, mostly, through the influence of the phenolic group upon the remainder of the molecule, resulting, in a large number of instances, in ring substitution para to the hydroxyl group.

In some cases the hydroxyl group is modified in the course of the reaction, as in the case of oxidation to quinone, the nitroso reaction to oxime, the production of urethans, and in alkylation or acetylation. In practically all of the other tests the hydroxyl group remains intact in the final product of the reaction, subject,

naturally in many cases, to the wandering of the labile hydrogen to produce tautomers. The oximes, produced by the nitroso reactions, are examples of the latter. (See second paper of this series.)

In cases where one or more of the hydroxyl groups remain in the final product, the acidic properties of this group determine the conditions under which color appears or changes of color take place.

It is to be noted that many of the tests enumerated for the phenolic group are produced with equal, or greater, facility by the amine group, in which case the reactions are usually much more complicated and quite apt to take place with the amine group directly.

The classification of many of the reactions involved in the phenol tests is simple; in some cases there is doubt regarding the exact character of the color formed, and in other cases no information is available upon which a logical classification can be based. I have endeavored to collect the important literature on the subject, and group the tests for purposes of simplicity and classification according to the chemistry of the reactions involved, wherever expedient.

A few of the tests have been satisfactorily applied on a quantitative basis for certain phenols; but the large majority is not susceptible of quantitative application, is not qualitatively specific for any phenol, and some of those that have been most employed by investigators are not specific for phenols as a class but are also given by many other substances not phenols. In employing such tests it is necessary that the investigator first be advised by some other means concerning the character of the material to be investigated. It is only then that many of the tests can be of any value and capable of intelligent application.

The very extensive literature treating of these tests is so widely scattered throughout chemical and other journals that much of it is not available to biologists. While the most of it is here presented no effort has been made to collect it all. However, it is believed that the references appended to this article together with those cited in the references will prove to comprise a fairly

complete list. Since an article on the indophenols is in preparation, which will include extensive references to these compounds, only the necessary references to this very interesting class of substances are here included.

In addition to the literature compilation the chemistry of the first group, the nitroso colors, has been elucidated by the work described in the second paper of this series. This group includes the many and complicated color tests employing dilute nitric acid, nitrous acid, alkyl nitrites and the various reagents of this class containing mercury salts, Millon's, Hoffmann's, Plugge's, Lintner's reagents and others.

In a later paper a new application of the indophenol test will be elucidated. It has been studied quantitatively and has been found to be very delicate for some phenols. This review is presented with the hope that it will be of assistance to chemists and biologists and also for the purpose of giving perspective to the experimental work now in progress and which will be presented in subsequent articles.

CLASSIFICATION OF TESTS

A. Dye reactions

A considerable number of the color reactions employed for the detection of phenols can be classified as producing dyes of one of the following six groups.

1. Nitroso colors
2. Nitro colors
3. Azo colors
4. Di- or triphenylmethane colors
5. Xanthene colors
6. Indophenol colors

These will be considered in the order named.

1. *Nitroso colors.* Nitrous acid reacts with many phenols with the production of compounds giving colored solutions of nitroso derivatives. In the presence of a condensing agent, such as sulfuric acid, the nitroso compound is condensed with the phenol giving the well known Liebermann, indophenol test.

It has been very difficult to differentiate the various tests where nitrous acid is employed and to classify them definitely. In the case of some indophenol tests both classes of colors are undoubtedly present, and in other cases condensation of the nitroso derivatives may take place in other directions.

In the case of certain tests, Nickel (1890), without experimental evidence, expressed the opinion that red dyes were formed from the nitrosophenols produced in the test, and in the case of salicylic acid Lintner (1900) holds the same view.

Tests employing dilute nitric acid are also described. It seems improbable under the conditions, that the nitric acid is present in sufficient concentration to cause nitration and it appears most probable that the color is due to nitrous acid present in the nitric acid or formed by the reducing action of organic compounds present.

It is interesting in this connection to note the action of nitrogen tetroxide and nitrogen pentoxide. Houston and Johnson (1925) found that the former acts as nitrous acid (nitrosyl nitrate) while Haines and Adkins (1925) found the latter to be an excellent nitrating agent.

The use of nitric acid mixed with sulfuric acid or hydrochloric acid is also described. In the case of sulfuric acid there is produced a nitrating mixture and this test is described under the nitro colors. In the case of hydrochloric acid very active chlorine is liberated and while there may be the effect of nitrous acid on the phenol this test is more fully treated under the halogen tests.

All of these reactions except those definitely belonging to the indophenol or nitro class are described under this heading. This classification is justified as a result of some of the experimental work described in the second paper of this series.

a. Colors formed by dilute solutions of nitrous acid, nitrites (salts of nitrous acid, alkyl nitrites, ethyl nitrite, amyl nitrite).

See previous discussion.

b. Colors formed by dilute solutions of nitric acid.

I have found that the use of the ordinary best reagent grade of nitric acid diluted, 3 drops in 10 cc. of water, will give a color

with a phenol solution, 2 parts in 1000, when an equal volume of the dilute acid is added. The color which appears on standing 10 or 15 minutes appears the same as that produced by dilute solutions of nitrous acid. (See second paper of this series.)

c. Colors formed by dilute solutions of nitric acid in presence of various salts (mercuric sulphate, etc.).

This test is similar to the preceding except for the catalytic action of the mercury salts.

d. Colors formed by nitric acid and nitrous acid in presence of salts of mercury and some other metals.

In this group is classed the very important and much used Millon's reagent (a nitric acid solution of mercury) made in various slightly modified ways, Hoffmann's reagent (a solution of mercuric oxide in nitric acid with the addition of a trace of nitrous acid), Plugge's reagent (a solution of mercurous nitrate with the addition of some free nitrous acid) and Lintner's reagent (a solution of mercuric nitrate, sodium nitrite, acidified with sulfuric acid).

See second paper of this series.

References

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|------------------------|-------------------------------|
| Millon (1849) | Perkin and Nierenstein (1905) |
| Hoffmann (1853) | Bach (1911) |
| Meyer (1864) | Elvove (1917) |
| Griessmayer (1871) | Weiss (1920) |
| Plugge (1872) | Chapin (1920) |
| Almén (1877) | Sen-Cupta (1921) |
| Allen (1878) | Salus (1922) |
| Hirschsohn (1881) | Huerre (1922) |
| Eykman (1882) | Ware (1925) |
| Fresenius (1883, 1889) | Houston and Johnson (1925) |
| Nickel (1890) | Haines and Adkins (1925) |
| Lintner (1900) | Ekkert (1925) |

2. Nitro colors. a. Nitration test. Many phenols can be nitrated to colored nitro derivatives. There is usually employed for this purpose a nitrating mixture of sulfuric and nitric acids. This reaction is one of the standard tests that has been used for a long time. In the case of phenol, picric acid results and by color comparisons of the acid or of its more highly colored salts with standards the test is made roughly quantitative.

References

- Mulliken (1914, pg. 108) Huerre (1922)
 Sen-Gupta (1921)

3. *Azo colors.* The Peter Griess diazo reaction and the coupling of the diazo derivative with phenols to form dyes has been found to be susceptible to development for its great delicacy as tests for phenols and for that reason has been extensively employed.

a. *Diazotized sulphanilic acid coupled with various phenols.* Ehrlich made use of diazotized sulphanilic acid and other diazotized amines in testing biological material but Hanke and Koessler first attempted to develop the test on a quantitative basis.

b. *Diazotized paranitraniline coupled with various phenols.* This test, as applied in a qualitative way, has been very popular but it has remained for Palkin and Wales to add a very valuable contribution in recording the spectroscopic investigations of 45 phenols most encountered in pharmaceutical preparations, which should render certain the identification of many of these compounds.

References

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|---------------------------------------|-------------------------------------|
| Ehrlich (1882, 1883, 1884) | Chapin (1920) |
| Penzoldt and Fischer (1883) | Fox and Guage (1920, 1922) |
| Nickel (1890) | Hanke and Koessler (1920, 1922) |
| Limpricht (1891) | Bell (1921) |
| Amann (1896) | Moir (1922) |
| Riegler (1899) | Ellms, Marshall and Phillips (1922) |
| Pauly (1904, 1905) | Theis and Benedict (1924) |
| Formanek (1905, 1908, 1913) | Palkin and Wales (1924, 1925) |
| Malacarne (1907) | Wales and Palkin (1926) |
| United States Pharmacopoeia, 9 (1916) | Pyman (1926) |
| Formanek and Knop (1917) | |

4. *Di- or triphenylmethane colors.* Various methods and reagents have been employed to perform this dye test. They produce brilliantly colored solutions of compounds which, in most cases, I believe should be classified in this group. The characteristics of the colors produced in many of the tests never have been sufficiently investigated, but from the mode of per-

forming the tests, dyes analogous to the triphenylmethanes will be produced.

In general the test consists in the condensation of an aldehyde with a phenol. A large number of aldehydes has been employed, such as formaldehyde, acetaldehyde, other alkylaldehydes, methyl glyococoll, vanillin, furfural, benzaldehyde, p-hydroxybenzaldehyde and others.

Other compounds producing reactions comparable to the aldehydes, such as phosgene, iodoform, bromoform, chloroform, chloral, carbon tetrachloride, oxalic acid, dihydroxyacetone, etc., have been employed.

As condensing agents sulfuric acid, hydrochloric acid, alkalies and other substances have been used.

The most generally employed reagents are formaldehyde and sulfuric acid.

a. *Marquis' reagent: Formaldehyde and sulfuric acid.*

b. *Ehrlich's reagent: Benzaldehyde and hydrochloric acid.*

c. *Guaraschi's test: Chloroform and alkali.*

References

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|----------------------------|------------------------------------|
| Kolbe (1861) | Lambert (1892) |
| Baeyer (1872) | Marquis (1896) |
| Guaraschi (1872) | Hehner (1896) |
| Michler (1876) | Barbet and Jandrier (1896) |
| Doebner (1878, 1879, 1880) | Endemann (1897) |
| Nencki and Schmid (1881) | Melzer (1898) |
| Sarauw (1882) | Deniges (1898, 1909, 1911) |
| Nencki and Sieber (1882) | Hartwich and Winckel (1904) |
| Lustgarten (1882) | Rhein (1907) |
| Ihl (1885, 1889) | Silbermann and Ozorovitz (1908) |
| Udránszky (1888) | Pouget (1909) |
| Schwarz (1888) | Sanchez (1911) |
| Crismer (1888) | United States Pharmacopoeia (1916, |
| Raupenstrauch (1888) | pg. 619) |
| Geigy (1889) | Sieburg (1916) |
| Nickel (1890) | Loele (1920) |
| Desesquelle (1890) | Salus (1922) |
| Caro (1892) | Gomberg and Snow (1925) |

5. *Xanthene colors.* a. *Phthalic anhydride fusion.* This test is essentially a fusion of phthalic anhydride with the phenol in

presence of a condensing agent. The phenolphthaleins long have been known but the colors of various derivatives were not systematically investigated until the work of Gsell, and Formanek and Knop. The former measured the absorption spectra in the visible region of 20 phenolphthaleins and the latter investigators presented a critical review of the work and also prepared some phenol anthraquinone derivatives.

Many phenols can undoubtedly be thus identified but this means of investigation is limited to cases where a considerable quantity, (e.g., 0.1 g.) of the phenol is available to carry out the fusion.

References

Baeyer (1876)
Mulliken (1914, pg. 107)
Gsell (1916)

Formanek and Knop (1917)
Orndorff and Murray (1917)

6. *Indophenol colors.* The name "indophenols" was given by Köchlin and Witt to a class of dyes developed by them about 1880; but long before this, about 40 years, colors were produced in various ways which we now recognize as due to indophenols.

These colors can be produced by various reagents applied in a variety of ways and the tests can be made very delicate. The acid solutions of the free indophenols are red while the solutions of the alkali and ammonium salts are intense blue.

It is interesting to note that Parry (1924) ascribes the color produced by Mandelin's reagent (1884), a vanadate in sulfuric acid, to the formation of a vanadium type of indophenol colors $O:C_6H_4:VH \cdot C_6H_4 \cdot OH$.

Previous to the work which will be described in a later paper of this series the tests have not been developed on anything but a qualitative basis. The new procedure depends upon indophenol formation which has been developed to a very delicate quantitative basis.

a. *Liebermann test.* Concentrated sulfuric acid containing nitrous acid. By this test the phenol is first partially converted to p-nitrosophenol which condenses through the agency of the sulfuric acid, with the phenol to form the indophenol.

Eijkman (1883) and Gutzkow (1889) employed amyl nitrite and sulfuric acid.

b. The simultaneous oxidation of an amine (p-aminophenol) and a phenol by a great variety of oxidizing agents will produce an indophenol. The oxidizing agent most commonly employed is hypochlorous acid.

c. Fluckiger test. This test depends upon the reaction of bromine upon the ammonium salt of a phenol. It produces an intense blue color and while it has not been investigated I believe it should be classed here and that the reaction is due to the bromine oxidation of the ammonium salt of the phenol rather than bromine substitution in the ring, although both may take place. It is well known that many oxidizing agents, including atmospheric air produce this reaction.

d. Quinonechloroimide tests. These tests employ various quinonechloroimides reacting with certain phenols to form indophenols and are fully treated in the experimental part of a later paper.

References

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| Berthelot (1859) | Hager (1884, 1885) |
| Lex (1870) | Waller (1887) |
| Neubauer (1872) | Wurster (1887) |
| Salkowski (1872) | Gutzkow (1889) |
| Jacquemin (1873) | Neubauer and Vogel (1890) |
| Rice (1873) | Manseau (1901) |
| Fluckiger (1873) | Kühl (1905) |
| Liebermann (1874) | Raschig (1907) |
| Cotton (1874) | Rodillon (1921) |
| Pollacci (1874) | Frieber (1921) |
| Almen (1877) | Parry (1921, 1924) |
| Eijkman (1883) | |

B. Halogen reactions

Halogenation tests have been applied extensively qualitatively and on a quantitative basis for detecting and estimating phenols. In many cases the methods are of considerable value in determining the identity of phenols, when sufficient quantities are available, since precipitates may be formed which can be purified and identified.

a. Iodination. Iodinated compounds of various phenols long

have been known and the methods of preparation are numerous; among which may be mentioned the use of iodine dissolved in alkali, iodine in potassium iodide solutions, iodine and mercuric oxide, iodine in sodium phosphate, iodine monochloride, potassium iodide and hypochlorite solution, and arylsulphonhalogenamides.

Cofman (1919) has shown that the active substance in the iodination of phenols is hypiodous acid, HOI, and that any method for the preparation of hypiodous acid can be used for the preparation of iodophenols.

As a quantitative method it has proved in most cases unsatisfactory although for some phenols satisfactory methods have been worked out, Emery (1921). See also Volumetric Iodate Methods, Jamieson (1926).

b. Bromination. Brominated derivatives have been employed both for the quantitative estimation and qualitative detection of phenols. As reagents bromates, bromites and free bromine have been employed.

c. Chlorination. The contamination of city water supplies by phenols from by-product coke ovens, and chemical works of various kinds has been evident by the objectionable odor and taste resulting from the chlorination of these waters either by hydrochlorite or free-chlorine according to water works practices. Comparisons of some of the various tests for phenols by Ellms, et al., lead them to the conclusion that the odor and taste tests are the most delicate for the purposes of water investigation.

The so-called euchlorine test employs potassium chlorate and concentrated hydrochloric acid. A reaction produced by a mixture of nitric and hydrochloric acids is classified under this heading since the reagent will generate chlorine. However, the test may well be attributed to the action of nitrous acid. The compounds formed have not been investigated.

Arylsulfonhalogenamides,—chloramine T (p-toluenedisulphochloroamide), halazone (p-sulfodichloraminobenzoic acid), and compounds of the type $\text{Ar}\cdot\text{SO}_2\text{NXM}$ where X = halogen and M = sodium or potassium, have been employed in halogenation, giving colors with various phenols.

Vitali (1892) describes a test employing a reagent consisting of concentrated sulfuric acid into which are put a few crystals of potassium chlorate. On adding a few drops of the solution to be tested there is first developed a green color changing to blue, if phenol is present.

I have tried this test and find that it is given by two drops of an 0.008 M (1:1,300) phenol solution. The mechanism of the reaction has not been investigated but it is natural to attribute it to chlorine dioxide. It may not be correct to list this test under the head of chlorinations but it is placed here for want of a better classification. If the sulfuric acid is contaminated with small quantities of oxides of nitrogen the indophenol reaction must be considered as a possibility.

References

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| Landolt (1871) | Wilkie (1911) |
| Koppeschaar (1876) | Hensel (1912) |
| Almén (1877) | Schewket (1913) |
| Allen (1878) | Mulliken (1914, pg. 109) |
| Hirschsohn (1881) | Folin and Dennis (1915) |
| Waller (1881) | Guglielmelli (1917) |
| Jacobson (1886) | Cofman (1919) |
| Vitali (1892) | Berthelot and Michel (1919) |
| Kossler and Penny (1893) | Desvernes (1920) |
| Neuberg (1899) | Emery (1921) |
| Kahn (1907) | MacLean and Thomas (1921) |
| Dehn and Scott (1908) | Ellms and Lawrence (1922) |
| Bougault (1908) | Heckner (1922) |
| Skirrow (1908) | Ellms, Marshall and Phillips (1922) |
| Mascarelli (1909) | Roberts (1923) |
| Mooser (1909) | Blaque (1923) |
| Neuberg and Hildesheimer (1910) | Vortmann (1923) |
| Liechti and Mooser (1911) | Jamieson (1926) |
| Seidell (1911) | |

C. Salts of metals

Under this heading there are classified a large number of tests alphabetically arranged according to the metal employed in the reagent. Attempts to work out the chemistry of the reactions employed have been recorded in the literature in only a few cases.

A most natural arrangement of many of these reagents would

place them definitely in the oxidizing class, the most obvious examples of which are, ferric chloride, dichromate, permanganate, etc. While such reagents undoubtedly act as oxidizing agents, the color formation is not to be explained so simply. Colored complexes may be formed with the metallic compounds, as has been definitely shown in the case of iron and titanium, and various condensations may take place with the oxidized phenols forming complex quinone derivatives. A number of the compounds of the metals entering into the reagents produce highly colored derivatives on reduction. In general it may be stated that in almost every case the color of the test is due to the reducing action of the phenol upon the salts of the metal or metals present and that the color of the oxidized phenol is not the predominating color of the reaction. It is therefore evident that in very few cases can these tests be made specific for any phenols and in fact the majority of them is undoubtedly not specific for phenols as a class, since any reducing substance will, in most cases, produce colors in the solutions of the reagents.

Antimony. A sulfuric acid solution of an oxide was employed by Levy (1886, 1887) to test a number of phenols and alkaloids. A number of other metals was also employed in the same way. See arsenic, bismuth, columbium, molybdenum, tantalum, tin, titanium and vanadium.

Arsenic. Angelo (1922) employed disodium arsenate (Na_2HAsO_4) and diammonium phosphate, $(\text{NH}_4)_2\text{HPO}_4$, either one producing a color when a small crystal is put into a 2 per cent phenol solution and then a small quantity of hydrogen peroxide added. The salts probably act as catalysts for the oxidation. See hydrogen peroxide test.

Guglielmelli (1916, 1917, 1918) devised tests employing arsenotungstic acid and arsenotungstomolybdic acid and compares the delicacy with 20 other of the known reagents. See also Lévy (1886, 1887).

Bismuth. See Levy (1886, 1887).

Cerium. Cerium sulphate gives a color with various phenols and it is stated that phenol can be made a very delicate test for cerium, 1 to 2000. Orlow (1907).

Chromium. Potassium dichromate has been employed as a reagent.

Cobalt. Bellucci and Chincini (1919) report that α -nitroso- β -naphthol gives a blood red color with cobalt compounds, is a very sensitive reagent and can be made the basis of a quantitative, colorimetric method for cobalt. Under certain conditions precipitates are obtained not only with cobalt but with other metals. Nickel (1888); Gugliamelli (1917).

Columbium. See Lévy (1886, 1887).

Gold. Auric chloride gives color reactions with phenols. Gugliamelli (1917); Fresenine (1883); Hirschsohn (1881).

The iron reagents. a. Ferric chloride. The blue color produced by ferric chloride reacting in solutions of many phenols has been a popular test for a long time. However, it is not very delicate. Sarauw (1882). Rashig (1907); Kahn (1907); Almén (1877).

Weinland and Binder (1912) describe certain phenol-iron complexes.

References

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|--------------------|------------------------------------|
| Waller (1902) | Hantzsch and Desch (1902) |
| Sarauw (1882) | Rashig (1907) |
| Hagar (1884) | Gibbs (1908) |
| Nickel (1890) | United States Pharmacopoeia (1910) |
| Bonanni (1900) | Wienland and Binder (1912) |
| Desmoulière (1902) | Mulliken (1914) |

a. Ferric alum. A solution of ferric alum is often employed in place of ferric chloride. United States Department Agriculture Bureau of Chemistry Bulletin No. 107 (1907).

b. Ferricyanide and ferric chloride. Jendrassik's (1923) reagent, consisting of potassium ferricyanide and ferric chloride, was found by Bezssonoff (1924) to be sensitive reagent for o- and p-phenols due to the reducing action of the phenols forming Prussian blue.

c. Ferricyanide and ammonia. This reagent has been employed by Candussio (1900). Pummerer et al. (1922, 1925) isolated a colorless ketone by oxidizing o-cresol with potassium ferricyanide in sodium carbonate solution.

d. Ferrous sulfate in presence of tartrates has been employed by Mitchell (1923), Price (1924) in the quantitative estimation of certain phenols found in galls and tannins.

Manganese. Potassium permanganate in alkaline solution will oxidize many phenols. Mulliken (1914, pg. 79.)

Mercury. Denige's reagent (1898), an acid solution of mercuric sulfate, has been employed by Escaich (1920) to distinguish between α - and β -naphthol, since α -naphthol gives a brilliant color while β -naphthol forms a yellow precipitate.

Millon's reagent, mercury dissolved in nitric acid, and other reagents of the same type containing mercury, are treated under the heading of the nitroso dyes.

Molybdenum. Fröhde's reagent is a solution of molybdic acid in sulfuric acid. Davy (1878), Allen (1878), Guglielmelli (1917), Lévy (1886). For the tungsten-molybdenum phosphoric acid complexes see tungsten.

Nickel. Color compounds of the type $\text{Ni}(\text{CN})_2 \cdot \text{NH}_3 \cdot \text{C}_6\text{H}_5 \cdot \text{OH} \cdot \text{H}_2\text{O}$ are described by Hofmann and Höchtlen (1903).

Phosphorous. Angelo (1922) employed diammonium phosphate and hydrogen peroxide in testing phenols. The former probably acts as a catalyst in the oxidation. See hydrogen peroxide test.

Platinum. Platinic chloride in some cases reacts somewhat similarly to gold. Fresenius (1883); Hirschsohn (1881).

Selenium. Levine (1920) states that selenious acid is a sensitive reagent for phenols and is of wide applicability.

Silver. Tollen's reagent, an ammoniacal solution of silver nitrate, is reduced by some phenols, especially many polyphenols. Mulliken (1914, fig. 22); Escaich (1920).

Sodium. Sodium peroxide has been employed as a test by Alvarez (1905).

Tantalum. See Lévy (1886, 1887).

Tin. See Lévy (1886, 1887).

Titanium. Hauser and Levite (1912, 1915) state that titanium salts form stable, colored complexes with phenols when two hydroxyl groups are present in the ortho position. An arseno-titanium reagent has been recommended by Guglielmelli (1917)

who also employed a sulphotitanium test. Also see Lévy (1886, 1887); Déniges (1916).

Tungsten. Tungstic acid has been employed in tests for phenols; however, the test most employed has made use of a reagent composed of tungsten and molybdenum complexes with phosphoric acid.

Reagents of this character prepared in many ways and of varying compositions, have been proposed as qualitative and, by color comparison with standards of known composition, quantitative methods for detecting and estimating phenols and many other compounds. This type of test has been extensively used by biologists and others.

Wu (1920) made a study of these complexes and it has been shown that the color reactions are produced by reduction. The reagent prepared in various ways is very sensitive to mild reducing influences. The color reactions are not to be attributed, or at least only in small part to the colored oxidation products of the phenols. The reagent is therefore not specific for phenols or in fact for anything except a reducing influence. Ever since Folin and Denis (1912) proposed this reagent it has been widely employed.

Under the heading of the arsenic reagents there is described Guglielmelli's modification.

References

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| Folin and Macallum (1912) | Scott (1921) |
| Folin and Denis (1912, 1915, 1916) | Bezssonoff (1921, 1922) |
| Johns and Jones (1918) | Levine and Burns (1922) |
| Benedict and Theis (1918) | Ellms, Marshall and Phillips (1922) |
| Folin and Wu (1919) | Rakestraw (1923) |
| Gortner and Holm (1920) | Henningsan (1923) |
| Wu (1920) | Goeffon and Nepveux (1923) |
| Levine (1920) | Cristol (1924) |
| Tisdall (1920) | Haas and Schlesinger (1924) |
| Chapman (1921) | Vorce (1925) |
| Donaldson (1921) | |

Uranium. Uranium salts form colored complexes with phenols. Siemssen (1912); Aloy and Laprade (1905); Aloy and Rabaut (1914).

Vanadium. Mandelin's reagent is a solution of a vanadate in concentrated sulfuric acid.

References

Mandelin (1884)
Lévy (1886)

Bellier (1899)
Parry (1924)

Zinc. Zinc chloride is employed as a condensing agent in producing the phenol phthalein colors. Zinc chloride and zinc sulfate are used by Nickel (1890) in connection with nitrous acid reagents.

D. Tests not classified in the foregoing

a. Acetylation. Acetylation of phenols has been employed in some cases for quantitative measurements. Verley and Bölsing (1902); van Urk (1921).

b. Alkaloidal precipitation. Hesse (1876) found that certain alkaloids formed precipitates with phenol and indicated that these might be employed as tests.

c. Urethan test. Herzog (1907) employed diphenylcarbonylchloride to produce diphenylurethans from phenols and French and Wirtel (1926) made use of alpha-naphthylisocyanate in the formation of urethans. A number of urethans were analyzed and characterized by their melting points.

d. Electrometric titration. Various phenols have been estimated by this method. Kolthoff (1920); Bishop, Kittredge and Hildebrand (1922).

e. Enzyme test. Bourquelot (1906) showed that *Russula delica* contained an enzyme that gave colors with a large number of phenols. Schmalfuss (1924) employed an enzyme from the blood of larvae of *acta caja* to differentiate between many phenols by means of the color formation. Onslow and Robinson (1925) found an enzyme in potato tubers that acted on phenols.

Mamas (1923) found a peroxidase in melanotic tumors that acted on polyphenols when two hydroxyl groups are in the ortho position to each other.

f. Esterification. Various esterification tests can be employed

to detect phenols. Hesse (1906) methylated isoborneol as a method of detection.

g. Hydrogen peroxide. Angelo (1922) obtained a fine rose or red color by oxidizing phenol with hydrogen peroxide in presence of diammonium phosphate or disodium arsenate. Without the addition of the salts the color development is much slower. For tests employing sodium peroxide see sodium. Wurster (1887); Kühl (1905).

h. Picrates. Goedike (1893) employed picric acid to precipitate various phenols; phenol, o-cresol, resorcinol, quinol, guaiacol and other.

i. Wood test. Runge test. This test depends upon the formation of a blue color in a splinter of wood (usually pine) on treatment of the wood first with phenol and then with concentrated hydrochloric acid. With some phenols the color is red. This test is severely condemned by Khotinskii.

References

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|----------------------------|----------------------------|
| Runge (1834) | Tommasi and Tommasi (1881) |
| Dispensatory, U. S. (1870) | Udránszky (1888) |
| Waller (1881) | Khotinskii (1917) |

DELICACY OF TESTS

Very few of the tests have any great degree of delicacy. The chlorine test as applied to potable waters, is claimed to be the most delicate and that even less than 1 part of phenol in 10,000,000 can be detected by the odor and taste.

Eykman claims 1:2,000,000 for the ethylnitrite test, and Wilkie 1:250,000 for the iodination test. The arsenotungstic, phosphotungstic-phosphomolybdic and the Millon reagents and the azo dye tests are stated to detect 1:1,000,000. The arylsulfonhalogenamides tests may be delicate in the case of β -naphthol, to 1:100,000 and the bromine test to 1:60,000. The ferric chloride test is not claimed to have a greater delicacy than 1:3,000 for phenol while for salicylic acid 1 part in 100,000 may be detected.

Various claims are made for some of the other tests, but it is

evident that the delicacy depends not only upon the conditions of the reagent and the solutions of the phenols, but also upon the character of the phenol present. The delicacy of a test that is more or less general for a number of phenols may vary greatly with the different phenols.

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